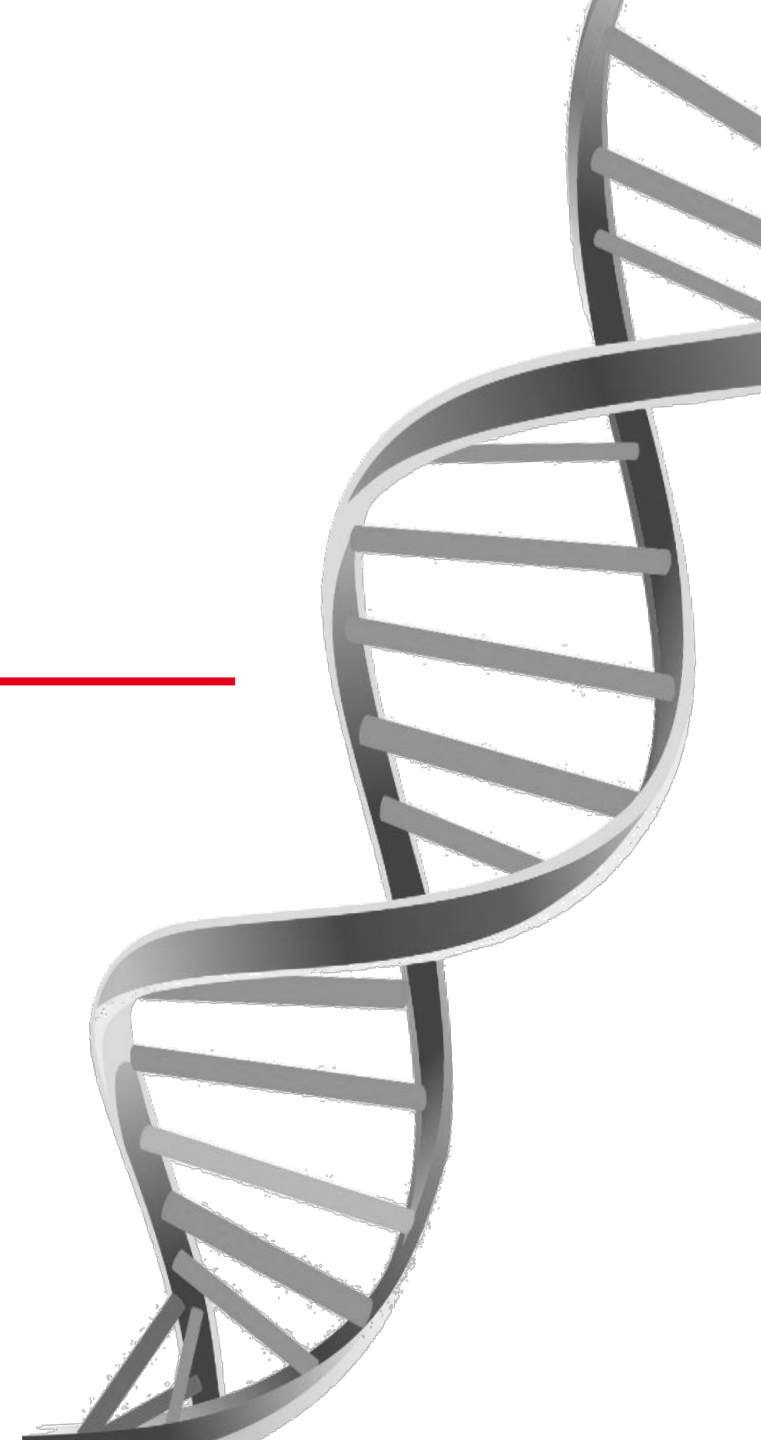




# Webinar #01

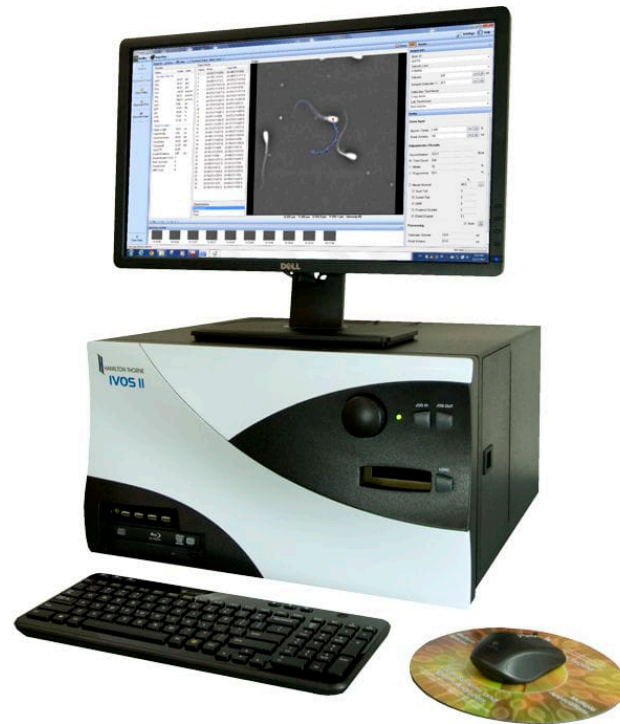
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Training department, v. 20200505





## WHY ASSESSING SPERM QUALITY ? WHY USING A CASA SYSTEM ?

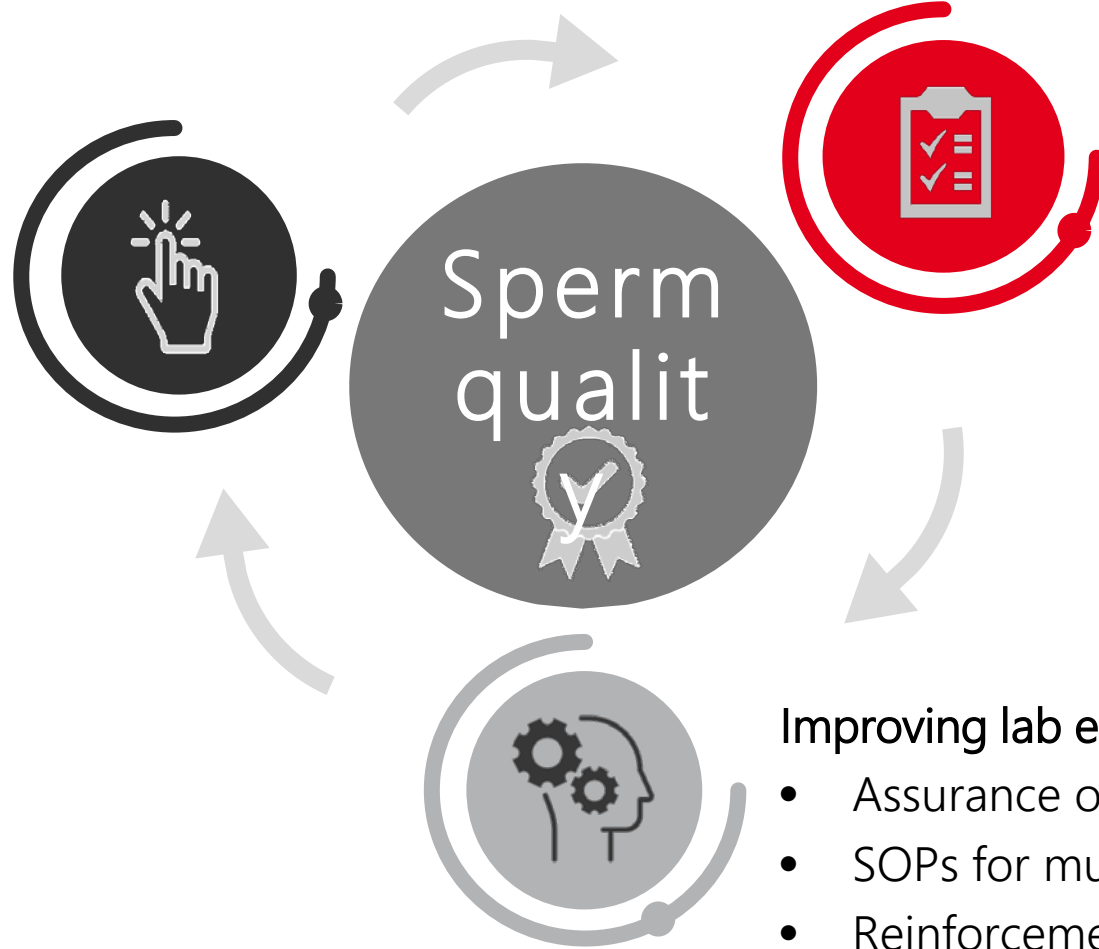


# Why assessing sperm quality ?

*Hossain et al., 2011; Amann et Waberski, 2014*

## Enhancing livestock management

- Decision tool to optimize sperm production and to better use high quality genetics
- Diagnosis of the health of the male reproductive organs
- Indication of environment conditions



## Standardized tool for protocols testing

- To improve sperm storage conditions
- To optimize fertility rates
- To enhance breeding/rearing management methods

## Improving lab efficiency

- Assurance of product quality
- SOPs for multi-sites management
- Reinforcement of the traceability
- To increase the number of doses produced thanks to a more accurate analysis of useful sperm



# Why assessing sperm quality ?

*Verstegen et al., 2002; Amann et Waberski, 2014; David et al., 2015*

## Sperm motility as an indicator of quality

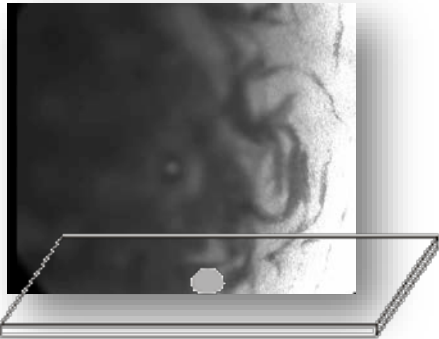
### Subjective motility



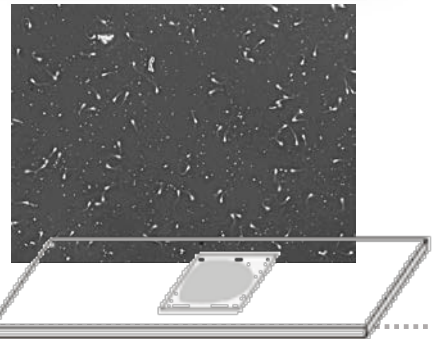
Use of a microscope and a human eye

- Time-consuming
- Operator-dependant
- Factors leading to mis-interpreting the results such as :
  - Sperm velocity
  - Sperm density
  - Sperm drift

Mass motility



Individual motility

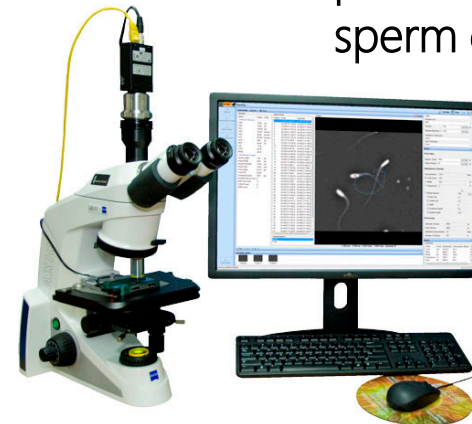


### Objective motility



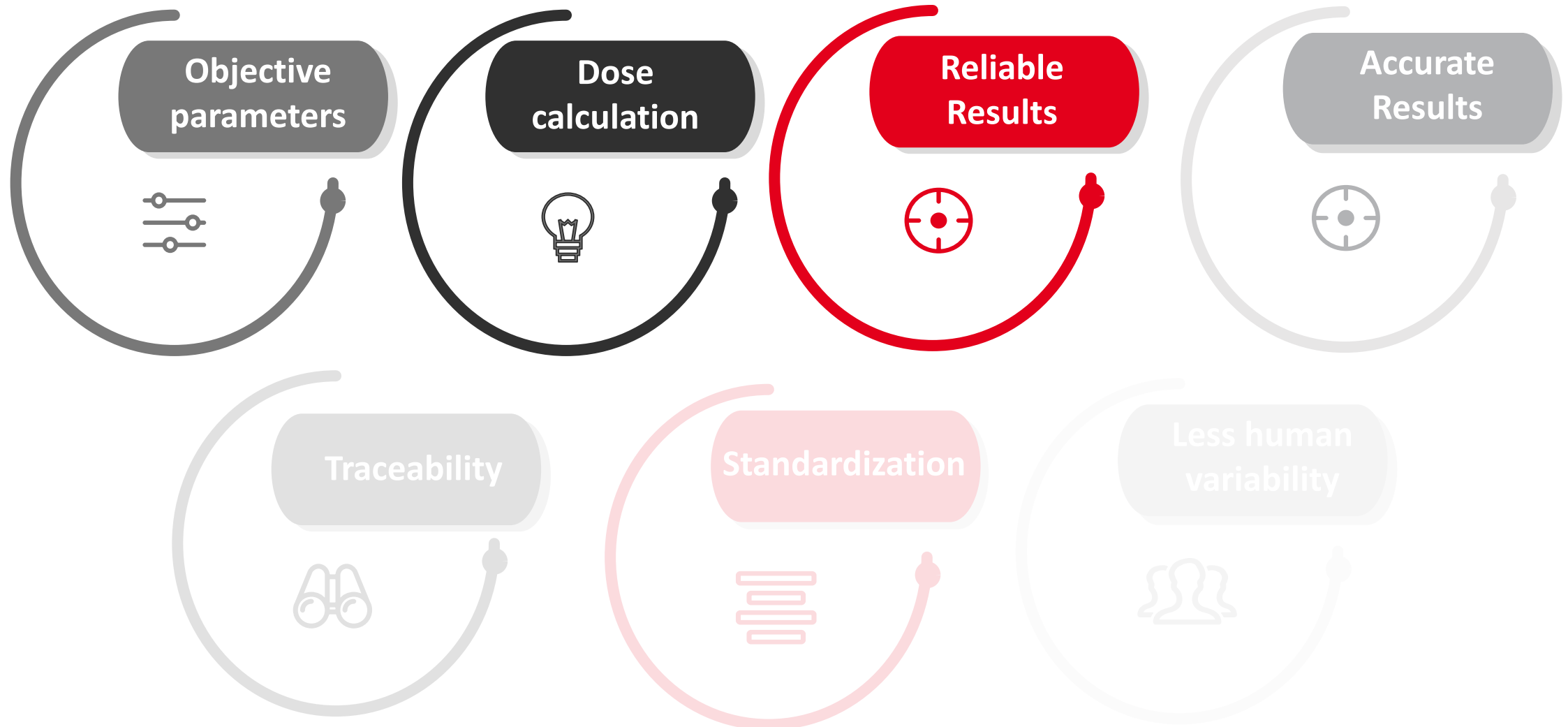
Use of a Computer-Assisted Sperm Analysis (CASA)

- Microscope linked to a camera
- A software able to quantify multiple motility & kinetics parameters of a high number of sperm cells, from a video



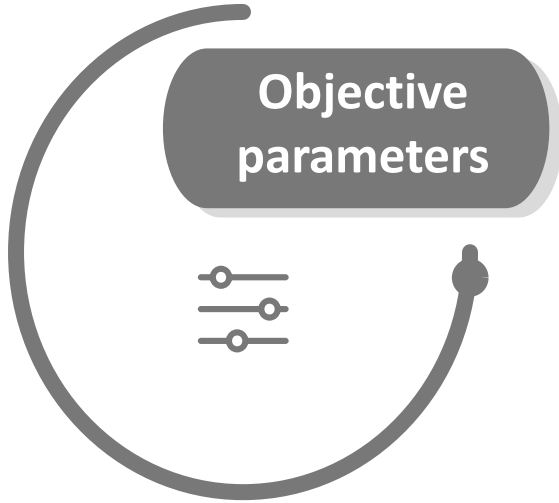
# Why using a CASA system ?

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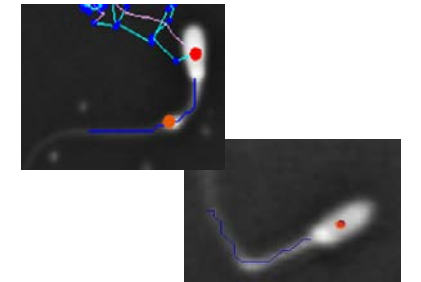
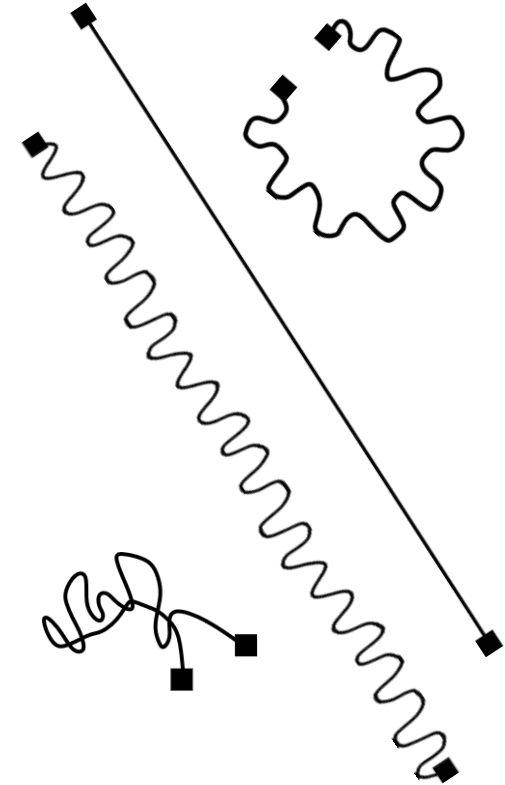
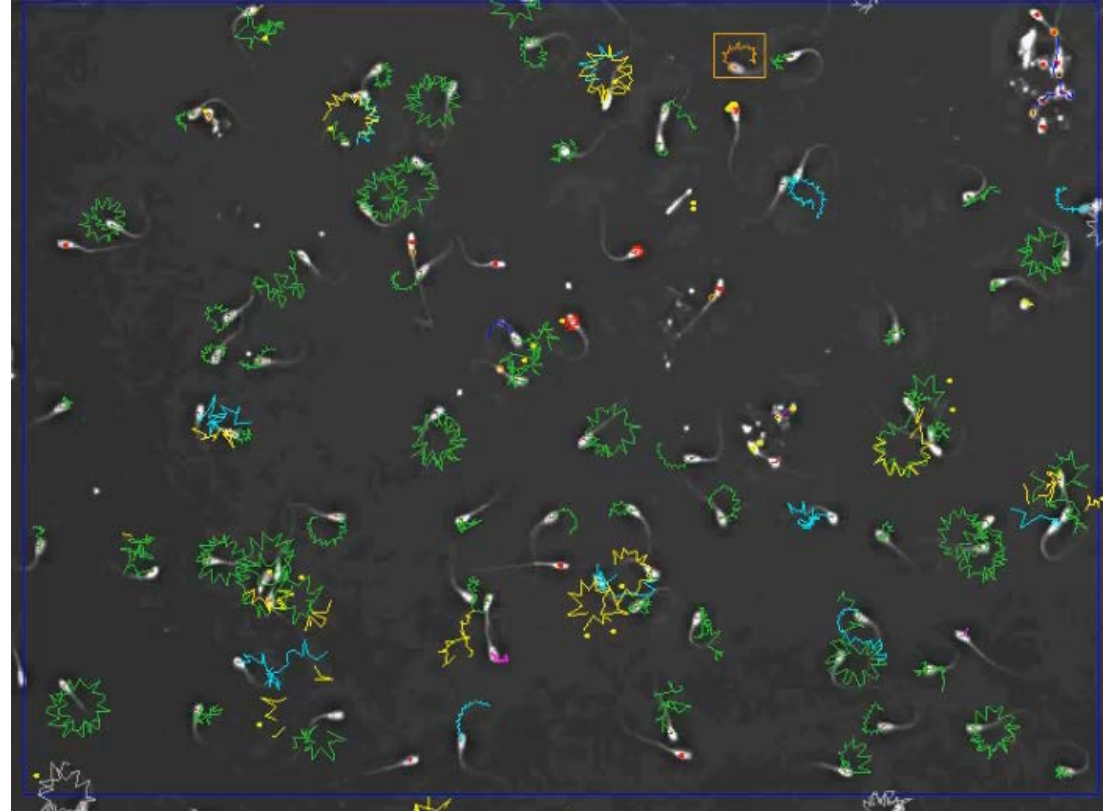




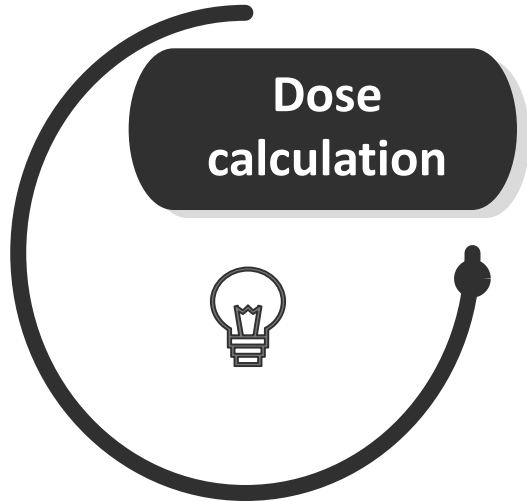
# Why using a CASA system ?



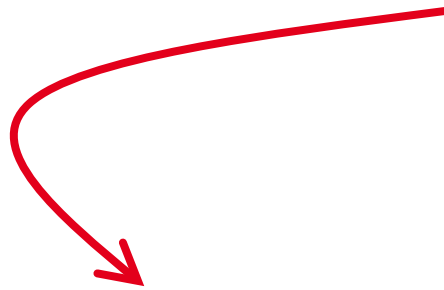
- Sperm tracking (motile/static)
- Kinematic (velocities, head movement)
- Concentration
- Morphologies



# Why using a CASA system ?

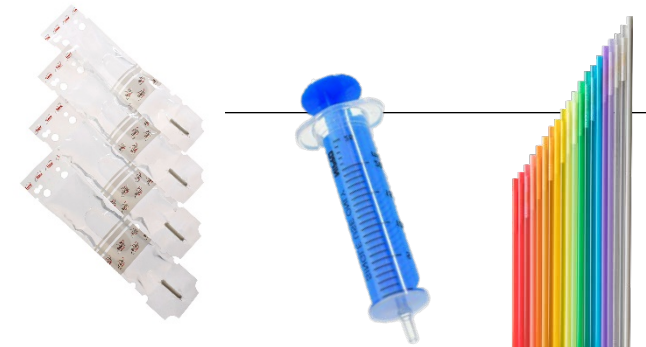


Input Your objectives	Output – Calculation by the computer
Initial concentration	Extender volume
Initial volume	Extender volume
Dose volume Sperm per dose Useful sperm per dose : % motile and/or normal morphology per dose	Dose number

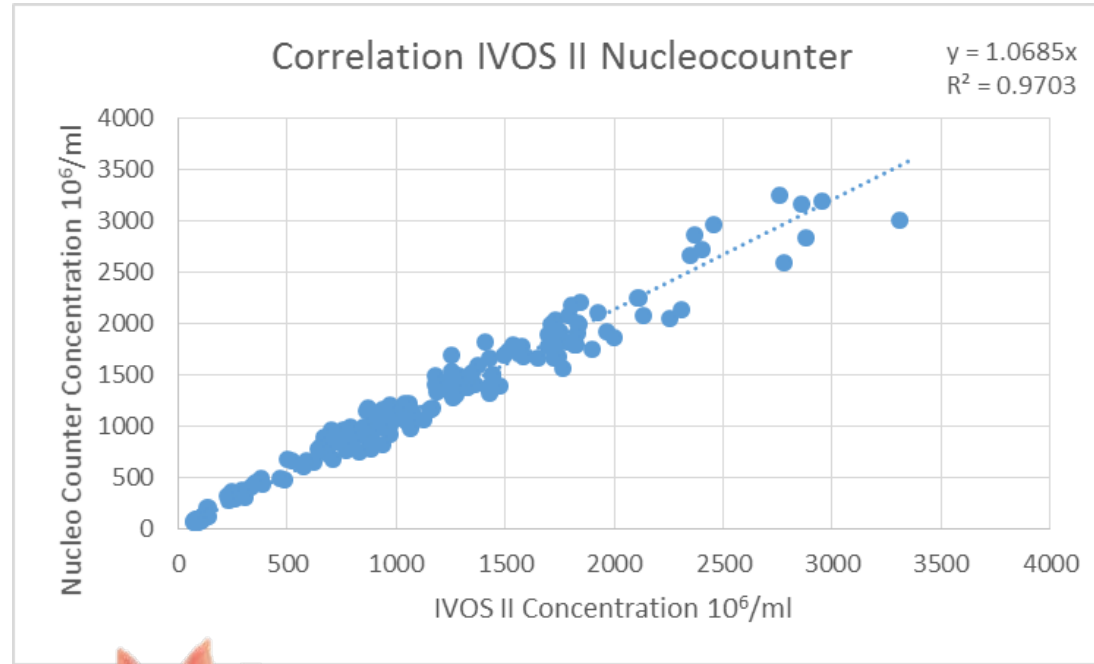
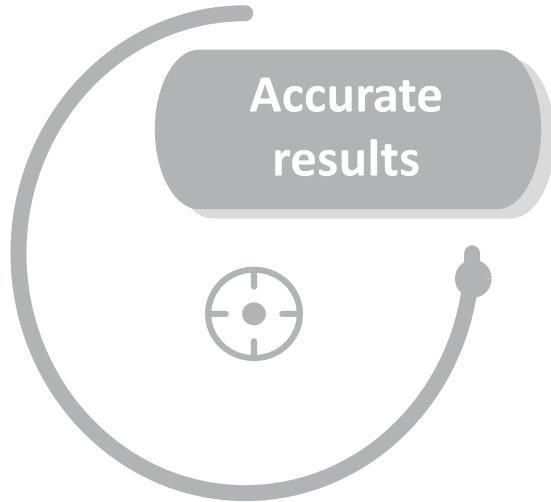


For an optimized stud production

*Dose packaging*



# Why using a CASA system ?



- Accuracy of parameters / results
- Very low CV = SD/Mean (< 10%)

	Pearson correlation coefficient	P	CV%
Concentration	0.96	P<0.0001	5.0
OMotility	0.98	P<0.0001	1.3
Morphology	0.98	P<0.0001	1.1



# Why using our IVOS II / CEROS II?

## Technical support and training provided



More than 30 years of expertise in sperm analysis



- Strong supplier collaboration
- High expertise in reproductive biotechnologies
- Large distribution in bull stud labs of a standardized setup in Europe (QualiVet, In Press)

High level training (2 days minimum)



- Mandatory for IVOS II & CEROS II installation

Technical & biological support



- Service contract for maintenance & re-training (anticipating team turnover)



# Why using our IVOS II / CEROS II?

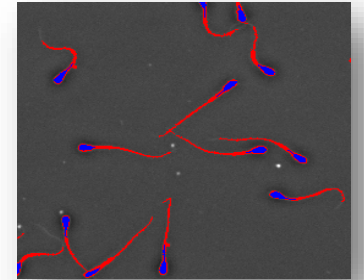
## Unique features of our IVOS II and CEROS II

**DEMO**



### Before analysis

- 'Live overlay' → heads/background differentiation
- Automated illumination LED regulation (IVOS II)
- Automated stage (IVOS II)

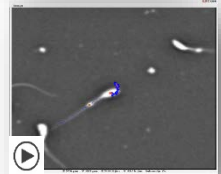
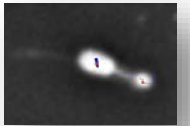


**Fast analysis !**

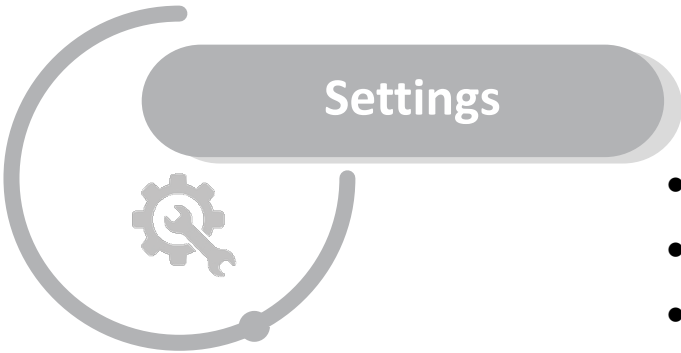


### During and after analysis

- Automated morphologies analysed, ie. DMR
- Playback of each field and of single spermatozoa
- Several digital image storage (mp4 + hmv)



**Intuitive software !**



### Settings

- Highly customizable setup
- Use of egg yolk-based medium possible
- Possible playback reanalysis with other setup



# Why using our IVOS II / CEROS II?

Mortimer & Maxwell, 2004



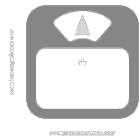
## Options available

### Fluorescence options



- Viadent® and Ident® = DNA target staining
- DNA Frag®

### Equipment

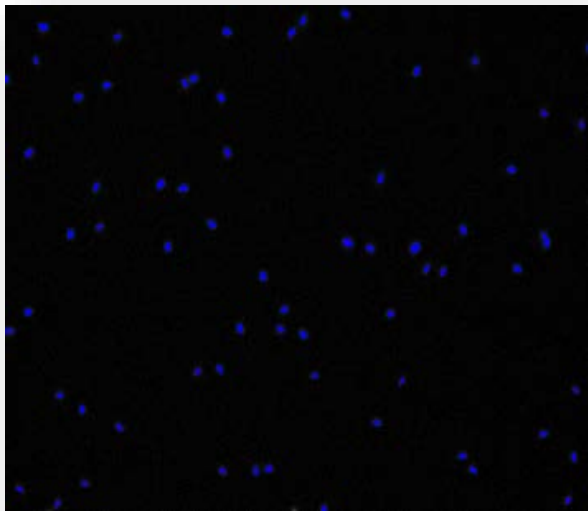


- Scale – volume input
- Footswitch – quicker analysis
- eSmile – stud management software

### Sort option



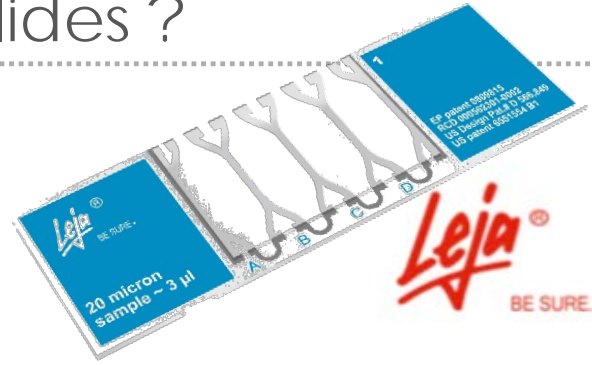
- Isolation of sub-populations of cells with customized setup
- Algorithm to detect hyperactivated sperm (Fractal Dimension, ie. FDM)



Dead sperm cells dyed with Viadent® on IVOS II



# Why using Leja slides ?



... Resulting in doses at accurate concentration !

# Why using Leja slides ?



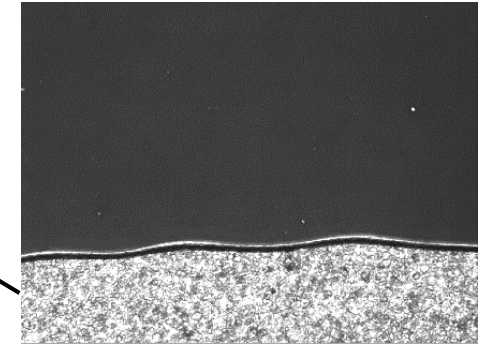
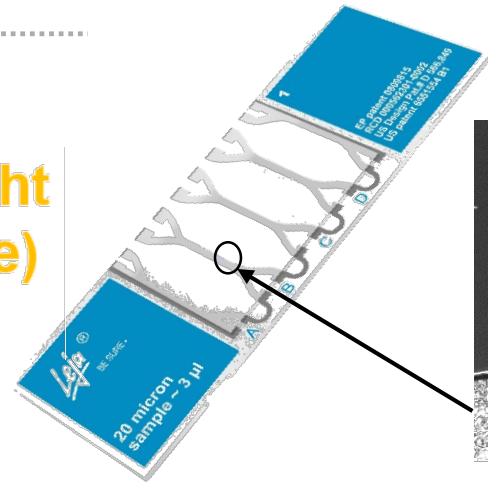
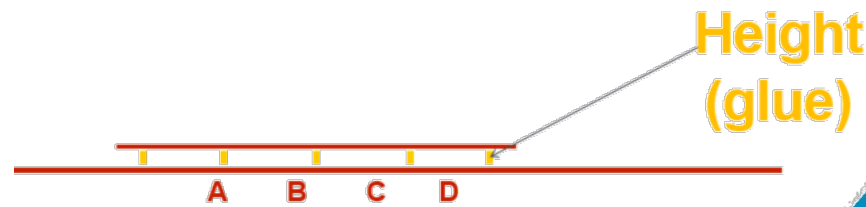
## ✓ Compared to slides and coverslips methods :

- Accurate concentration estimation
- No air bubbles or agglomerates
- Homogeneous repartition (no rheotaxis effect)
- No risk of looking for the best frames and then misinterpreting results
- **Constant height...**

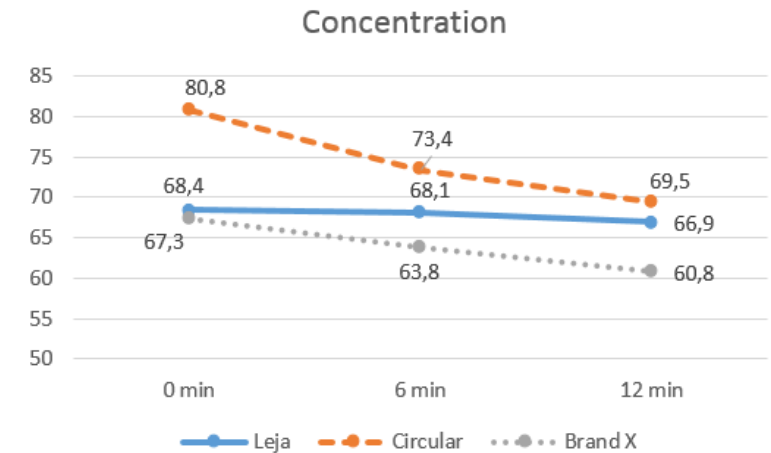
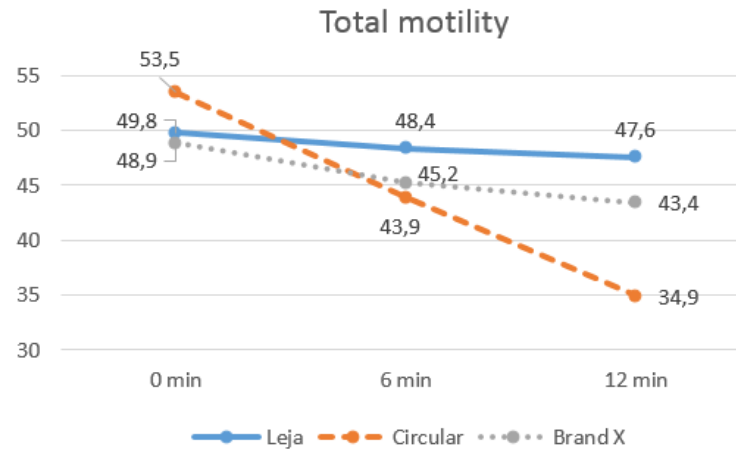


# Why using Leja slides ?

Ibanescu et al., 2016



- High quality glue resulting in :
  - Fixed depth counting chamber
  - No coating marks
  - No air bubble development
  - No sperm toxicity
  - Accurate concentration estimation



Total sperm motility and concentration according to the chamber type and time after filling (n = 60).

Adapted from Ibanescu et al., 2016

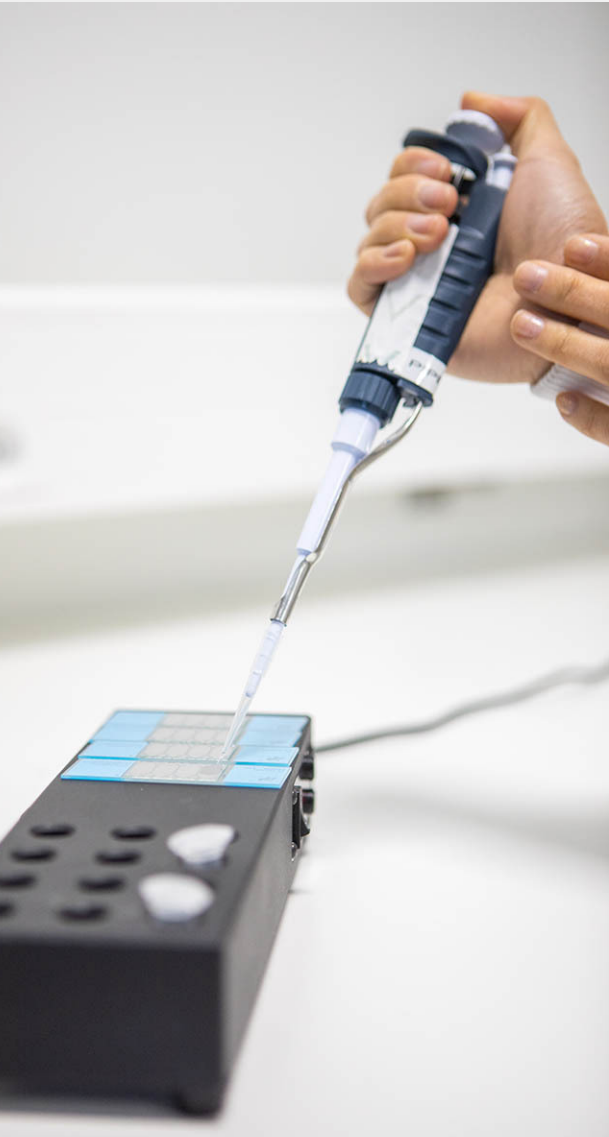




# Why using our range of sperm quality assessment ?

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## Conclusion



Accurate and reliable  
control quality of sperm

Better use of **high quality genetics**

**High standardization** of your lab procedures

**Entire range** for sperm analyses for an **enhanced efficiency at the lab**  
(Associated equipment, **Leja<sup>®</sup> slides**, eSmile, etc.)

**High level** of technical and **biological support**

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Thank you for your attention

**imv**  
TECHNOLOGIES

Questions ?

# References

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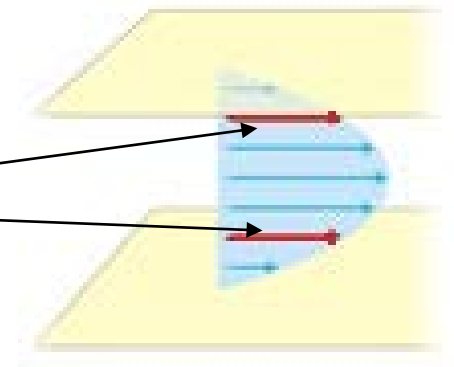
## SERGE SILBERBERG EFFECT

The distance of these planes from the wall ( $\beta$ ) is depending on a few parameters:

- Development of full Poiseuille flow
- Chamber height
- Surface properties of the counting chamber
- Surface tension
- Fluid viscosity
- Flow velocity
- Diameter of sperm-cell head

Sperm cells in the middle of the chamber-height **move faster** than the ones near the wall

Sperm cells move to two planes at equidistance from each chamber wall



References (to download at [www.leja.nl](http://www.leja.nl)):

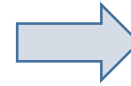
Douglas-Hamilton, D.H.; Smith, NG; Kuster, C.E.; Vermeiden, J.P.W.; Althouse, G.C.: Particle Distribution in Low-Volume Capillary-Loaded Chambers, *Journal of Andrology*. 2005;26(1):107-114

Douglas-Hamilton, D.H.; Smith, NG; Kuster, C.E.; Vermeiden, J.P.W.; Althouse, G.C.:Capillary-Loaded Particle Fluid Dynamics: Effect on Estimation of Sperm Concentration, *Journal of Andrology*. 2005;26(1):115-122

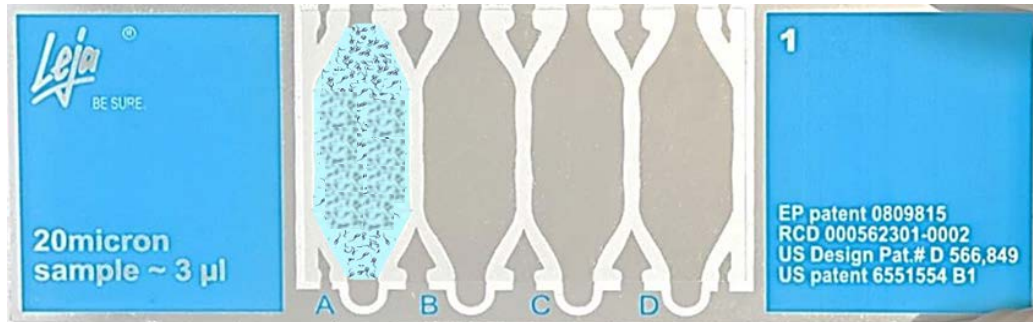


## SERGE SILBERBERG EFFECT

Sperm cells in the 2 Segre Silberberg planes move faster than the average fluid velocity



Accumulation of sperm cells at the filling front.



When measuring the sperm concentration in the center of the Leja slide, an **underestimation of the concentration** takes place.

This is a **constant underestimation** that can be **corrected!**

References (to download at [www.leja.nl](http://www.leja.nl)):

Douglas-Hamilton, D.H.; Smith, N.G.; Kuster, C.E.; Vermeiden, J.P.W.; Althouse, G.C.: Particle Distribution in Low-Volume Capillary-Loaded Chambers, *Journal of Andrology*. 2005;26(1):107-114

Douglas-Hamilton, D.H.; Smith, N.G.; Kuster, C.E.; Vermeiden, J.P.W.; Althouse, G.C.: Capillary-Loaded Particle Fluid Dynamics: Effect on Estimation of Sperm Concentration, *Journal of Andrology*. 2005;26(1):115-122







## SERGE SILBERBERG EFFECT

<b>Cell Detection</b>	
Elongation Max (%)	60
Elongation Min (%)	5
Head Brightness Min	123
Head Size Max ( $\mu\text{m}^2$ )	70
Head Size Min ( $\mu\text{m}^2$ )	14
Static Tail Filter	False
Tail Brightness Min	82
Tail Min Brightness Auto Offset	10
Tail Min Brightness Mode	Auto - First Frame
<b>Chamber</b>	
Capillary Correction	1.5
Chamber Depth ( $\mu\text{m}$ )	19.6
Chamber Type	Capillary
<b>Illumination</b>	
Intensity Visible	2244
Max Photometer	90
Min Photometer	49
<b>Kinematics</b>	
Progressive STR (%)	75
Progressive VAP ( $\mu\text{m/s}$ )	60
Slow VAP ( $\mu\text{m/s}$ )	30
Slow VSL ( $\mu\text{m/s}$ )	20
Static VAP ( $\mu\text{m/s}$ )	2
Static VSL ( $\mu\text{m/s}$ )	1
<b>Morph</b>	
DMR Confidence (%)	10
DMR Droplet to tail end Max ( $\mu$ )	5

(Name)  
User definable setup name

OK Cancel Apply

This Sildeberg effect is taken into account in the software

Chamber Depth has to be correct

**Dosing**

**Dose Input**

Sperm / Dose	2.500
Dose Volume	80.00
Usable Volume	80.00

**Adjustments / Results**

Concentration	B/ml
<input checked="" type="radio"/> Total Count	
<input type="radio"/> Motile	%
<input type="radio"/> Progressive	%
<input checked="" type="checkbox"/> Morph Normal	%
<input checked="" type="checkbox"/> Bent Tail	
<input checked="" type="checkbox"/> Coiled Tail	
<input checked="" type="checkbox"/> DMR	
<input checked="" type="checkbox"/> Proximal Droplet	
<input checked="" type="checkbox"/> Distal Droplet	

**Processing**

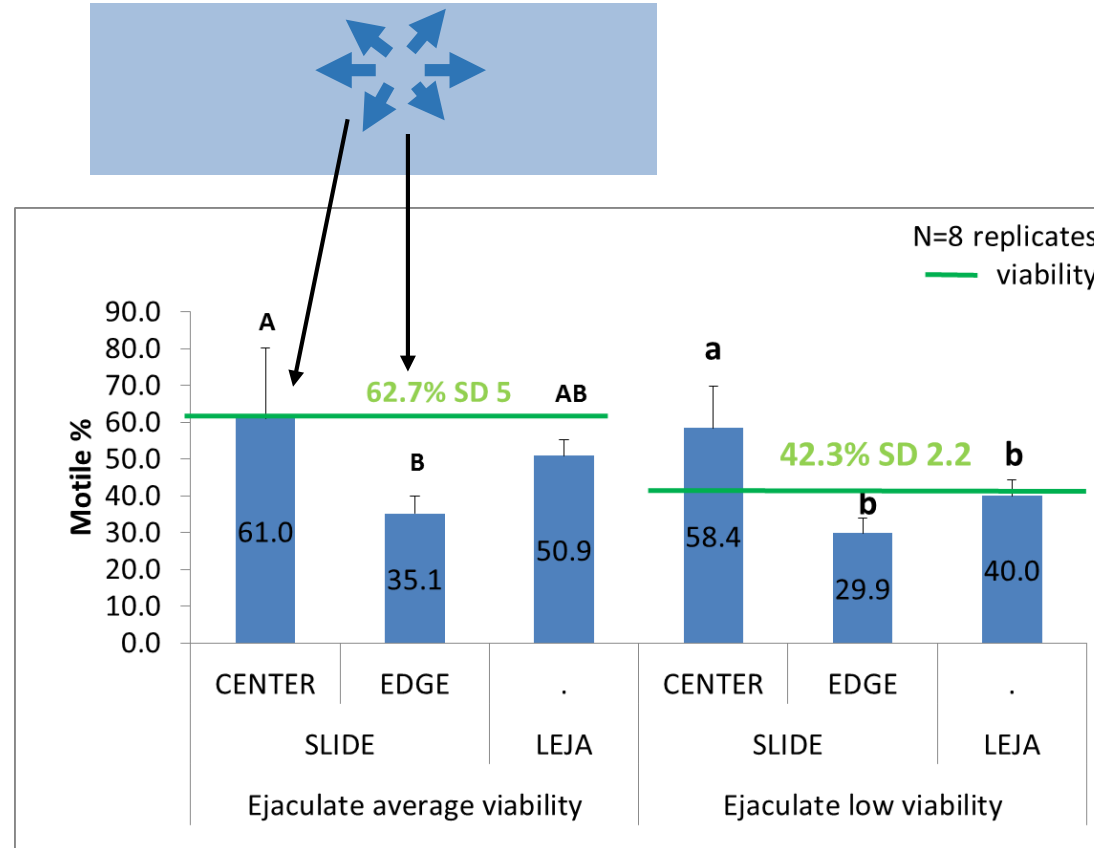
Extender Volume		ml
Final Volume		ml
Adjusted Concentration		B/ml
Number Of Doses		#

IVOS II will calculate Concentration according to these parameters





# Sample loading



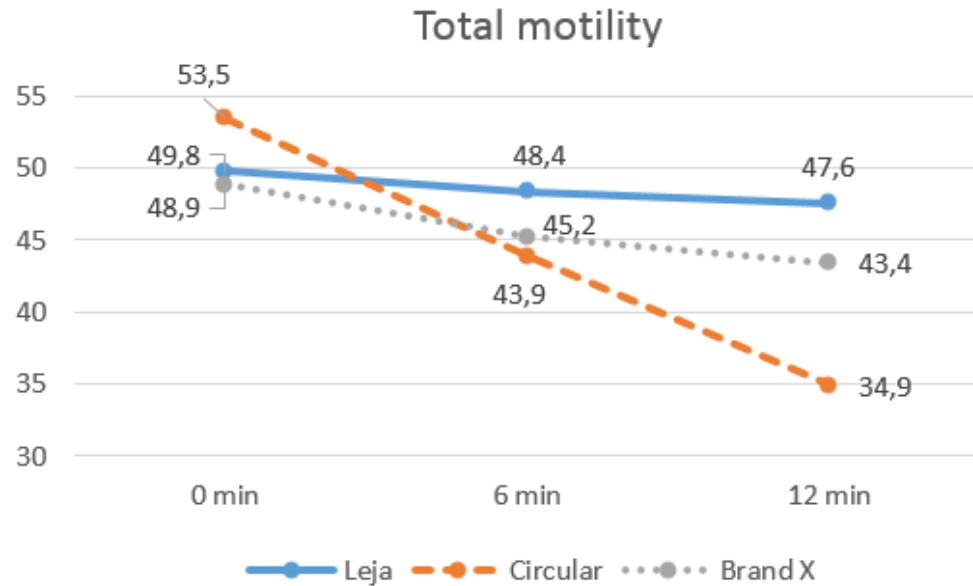
Slide + Coverslip leads to **misinterpretation**

Leja® gives more precise and repeatable results

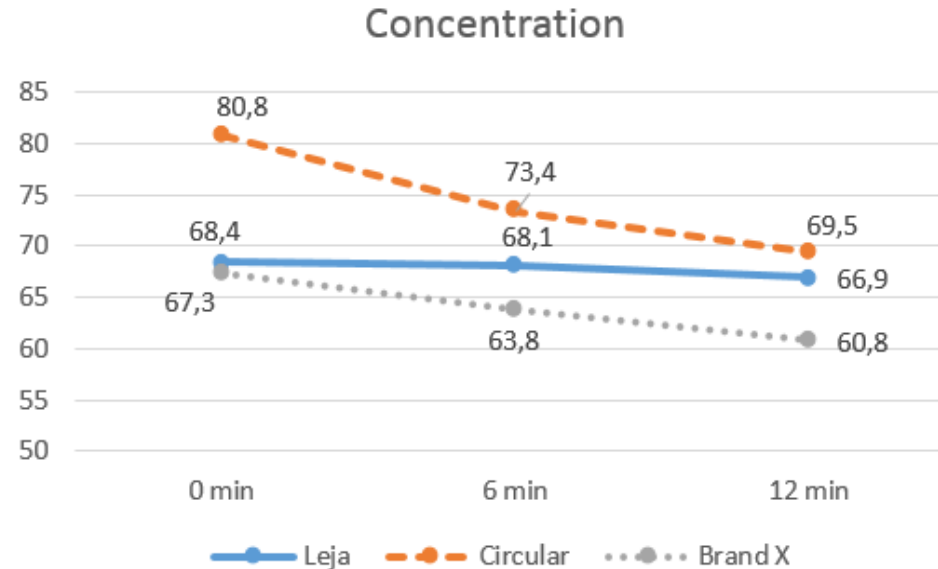


# Sample loading

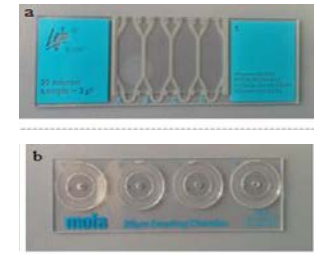
Ibanescu et al., 2016



Total sperm motility according to the chamber type and time after filling (n = 60).



Sperm concentration according to chamber type and time after filling (n = 60).



“Reliable results (motility and conc.) for Leja after 6 and 12 min ( $p < 0.05$ )”

**Circular and Brand X effect of toxicity over the time and sperm distribution**



True values	
<u>Boar ejaculate</u>	
Volume:	300 ml
Concentration:	300 million cells/ml
Dose concentration	1.5 billion cells/80 ml
True motility	75 %
results in 45 doses	

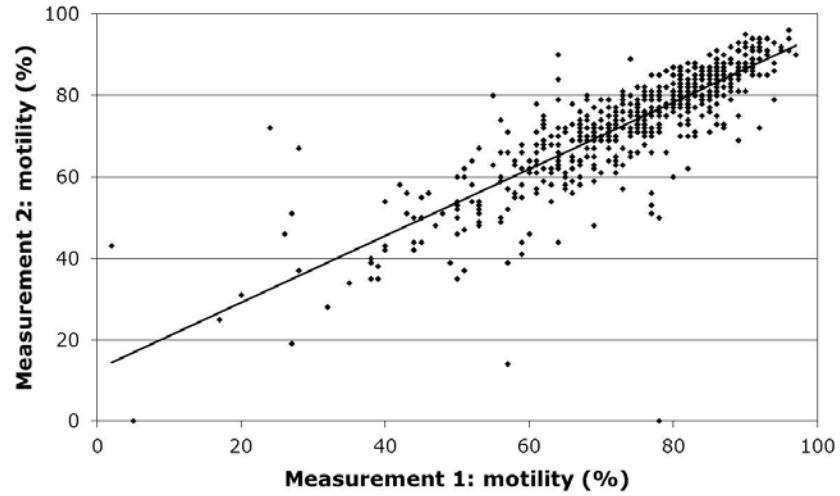
Low repeatability (without training)		High repeatability (trained and with SOP)	
Repeatability = 71% Motility min 53 % max 97 % difference ± 22 % ↓		Repeatability = 96% Motility min 72 % max 78 % difference ± 3 % ↓	
Doses produced min 32 max 58 difference ± 13		Doses produced min 43 max 47 difference ± 2	
58 R I S K 45 45 L O S S 32		47 45 43	

Low repeatability consequence :

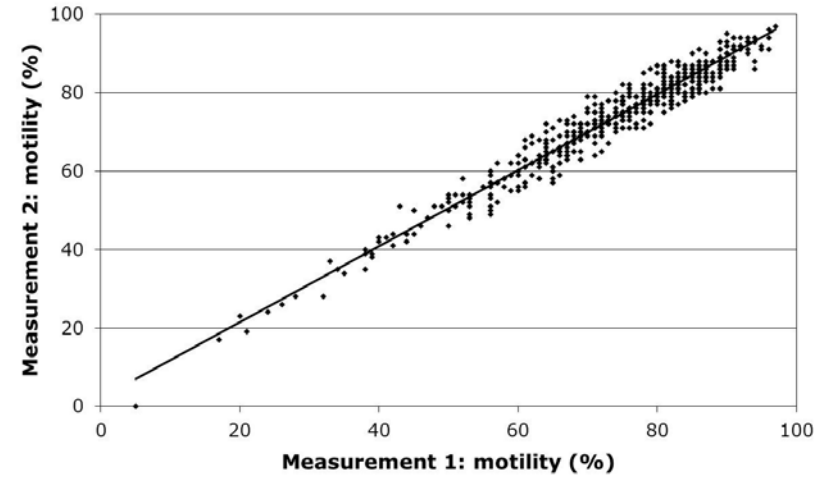
overestimated values for the number of doses  
 ⇒ risk of insufficient fertility (insufficient concentration)

underestimated values for the number of doses  
 ⇒ risk economic losses for the center (insufficient number of doses produced)

*Broekhuijse et al., 2011*



Before training



After training

